Osteoarthritis (OA) of the knee is the most common type of arthritis, and as such, it is the main cause of pain and disability in the elderly [1,2]. The risk factors related to OA are numerous and include past joint trauma or repeated micro-trauma, advanced age, obesity, nutritional factors, female gender, occupation, joint deformities, muscle weakness and genetics [3]. Out of all patients diagnosed with OA, approximately 12% have clear evidence of trauma prior to the onset of symptomatic disease [4]. The avascular nature of articular cartilage limits its regenerative and healing capacity; therefore, patients with post-traumatic OA (PTOA) heavily depend on external treatments designed to minimize damage to the joint structures and support the healing process. There is also a clear need to reestablish joint functionality in these patients because mechanical loads are critical to maintaining the health of the articular cartilage, bone, and muscles [5,6]. The pain caused by trauma, damaged tissue, and subsequent inflammation often creates a vicious cycle that starts with decreased physical activity and leads to atrophy of the joint and ultimately, physical debilitation [7,8]. Ample preclinical and clinical evidence suggests that physiologic loads are not only beneficial but necessary for the health of the joint [9-11]. The incapacity of the osteoarthritic limb to withstand physiologic loads is a critical element in OA pathophysiology; thus, the reestablishment of mechanical function through moderate exercise is recommended for the management of OA patients [12,13]. Because no effective therapies that specifically target chondrocytes exist, clinical guidelines recommend both pharmacological and non-pharmacological approaches to relieve the symptoms of OA [14]. Both bone and cartilage turnover are deemed critical for the health of synovial joints, hence the use of both antiresorptive and anabolic bone agents could be used to treat OA symptoms [15,16]. Preclinical data overwhelmingly indicate that antiresorptive [17,18] and anabolic [19,20] therapies curb progression of OA in animal models, however, clinical results fail to demonstrate clear benefits of these therapies to OA patients despite the fact that these drugs have been in clinical use for many years [21,22].

A unilateral medial meniscectomy (MM) in rats results in the progressive degeneration of the articular cartilage with subsequent sclerosis and osteophyte formation, which further limit joint function [23]. The recent study conducted by our group showed no significant beneficial effects of either antiresorptive or anabolic therapy on the key denominators of OA, including cartilage damage, osteophyte formation, osteosclerosis and joint functionality [24]. Although our study results aligned with the clinical data, we did not expect a “no-effect” outcome, particularly because the vast majority of published preclinical data shows that bone-targeting drugs have at least moderate efficacy in animal models of OA [17-20]. To further elucidate the possible reasons for the abovementioned discrepancy...
and to further extrapolate the ability of bone-targeted therapy to improve joint functionality and bone strength, we evaluated the cortical bone geometry and cortical bone strength of the femurs of injured (underloaded) and contralateral legs in a rat model of PTOA.

**Methods**

**Animals and management**

Four-month-old male Lewis rats (Charles River Laboratories, Portage, MI, USA) weighing 350 g were used in this study. All in vivo procedures were approved by the Institutional Animal Care and Use Committee (IACUC) at Pfizer and were performed in accordance with the Guidance [25]. The rats were pair-housed in ventilated cages (Innovive Inc., San Diego, CA, USA) in a temperature- and humidity-controlled room on a regular 12-hour light/dark cycle. Irradiated LabDiet® 5053 (Purina, Richmond, IN, USA) and water were provided *ad libitum*. The rats were acclimated for one week prior to their use in the study. A total of 48 rats were used for the 10-week study, with 12 rats per group. One group of 12 rats received sham surgery; the remaining 36 rats underwent MM surgery. The four study groups were as follows: a sham control (Sham), an MM vehicle-treated (MM/Veh), an MM + zoledronate (MM/Zol) and an MM + PTH (MM/PTH).

**Surgery**

The rats were anesthetized using isofluran (Pfizer Animal Health, New York, NY, USA) and sustained-release buprenorphine (Zoopharm, Windsor, CO, USA) were administered prior to surgery for analgesic coverage. In the sham group, a surgical approach to the medial collateral ligament on the right hind limb was completed by cutting the skin and leaving the medial collateral ligament intact. In the surgery groups, MM was performed by fully transecting both the medial collateral ligament and the medial meniscus of the right hind limb, followed by closure in 2 layers using absorbable sutures [23].

**Dosing and bone labeling**

The rats in the sham and MM/Veh control groups received vehicle (sterile water) at 1 mL/kg subcutaneously (sc) 5 days/week starting on the day of surgery. The rats in the MM/Zol group received zoledronic acid (Zol; Sargent Pharmaceuticals 25021-801, Schaumberg, IL, USA) at 100 µg/kg sc twice/week [16], and the rats in the MM/PTH group received human PTH (bPTH 1-34; Sigma-Aldrich, P3796, St. Louis, MO, USA) at 40 µg/kg sc five times/week [20] starting on the day of surgery. To label the actively mineralized bone surfaces calcine (Sigma-Aldrich Cat# C-0875) at 10 mg/kg (3.3 mL/kg) was injected 13 days before necropsy and alizarin (Sigma-Aldrich Cat# A-5533) at 30 mg/kg (3.0 mL/kg) 3 days before necropsy.

**Body weight, sample collection**

Body weight was recorded twice weekly throughout the study. At the end of the 10-week study, both hind legs were carefully harvested and wrapped in saline-soaked gauze and frozen at -20°C for *ex vivo* imaging, mechanical testing and histological analyses.

**Dynamic weight bearing**

Dynamic weight-bearing (DWB) measurements were obtained before surgery, at week 5 and before euthanasia to assess the effects of surgery on the weight-bearing capacity of the hind and front legs using a BIO-SWB-R model (Bioseb, Boulogne, France, version 1.3) and method described earlier [26].

**Radiology**

All of the knee samples were X-rayed with a Faxitron Model MX20 specimen scanner (Faxitron Biopics LLC., Tucson, AZ, USA) using exposure time of 12-18 sec at 31-35 kV and 3 x magnifications to inspect the bone samples possible abnormalities.

**μCT and EPIC μCT measurements**

Pre-contrast scans of all the tibias were obtained using the MicroCT 100® computed tomography system (Scanco Medical, Bassersdorf, Switzerland) with the following parameters: 800 slices, 10-µm resolution, a total scanned area of 8.0 mm², and source energy of 70 kVp, 115 µA at 8 W to capture the entire proximal tibia section. The tibias were then incubated in the Hexabrix solution and scanned using a previously described method [24,27]. Post-soak scanning of the right tibia was performed using source energy of 55 kVp, 145 µA at 8 W and an average scan time of 42 min per sample.

**μCT evaluation of the epiphyseal cartilage**

Using the post-contrast scans, contour lines were drawn around a range of interest (ROI) that included the cartilage overlying the medial tibial plateau as described earlier [24,28]. Other ROIs were drawn and analyzed on this central midpoint of the articular surface in a manner corresponding to the standard histological evaluation techniques for the articular cartilage [29]. The length of the medial articular cartilage was measured and divided into 3 zones of equal length, and the cartilage volume was evaluated as described elsewhere [24].

**Cartilage histology**

After the EPIC μCT imaging of the articular cartilage was completed, six tibias were randomly chosen and placed in 10% neutral buffered formalin for 72 h prior to demineralization for 8 days in Immucal (Decal Chemical Corp. Tillman, NY, USA). The tibias were then processed in paraffin and serially sectioned at ~200-µm intervals into 5-µm-thick sections for with hematoxylin and eosin, toluidine blue and safranin O to evaluate cartilage damage using the method suggested in the literature [24,29,30]. Here, we report the overall histology findings and zonal analysis of the cartilage thickness parameter, as evaluated using toluidine blue sections.

**μCT evaluation of the cortical bone at the femoral mid-diaphysis**

μCT of the cortical bone mid-diaphysis was conducted on both the left and right femurs using a μCT-100® computed tomography system and previously described method [31]. Sample scans were performed on 25 slices (1 slice = 10.5 µm) using high-resolution settings. The
following parameters were evaluated bone mineral density, tissue volume, bone volume, bone marrow volume, cortical thickness, bone area, polar moment of inertia, maximal I value, minimal I value and $I_{\text{MAX}}/C_{\text{MAX}}$.

**Bone strength testing using the 3-point bending method**

Both the left and right femurs were mechanically tested with using an Instron materials testing machine (5543A, Instron Inc., Norwood, MA, USA). The femurs were positioned cranial side up across two lower contacts that had a span of 5-7 mm, with an upper contact centered between the lower contacts. The bone was broken in 3-point bending using a cross-head speed of 0.5 mm/min. During testing, force and displacement data were collected at a frequency of 200 Hz using the BlueHill 3 testing software, version 3.41 (Instron Inc). Force/displacement curves were generated, and ultimate force (maximum load), stiffness (maximum slope) and energy to fracture were recorded.

**Dynamic histomorphometry of the cortical bone**

The distal ends of the femoral cortical bone from the left and right femurs were collected after mechanical testing and embedded undecalcified in methylmethacrylate, cut into 20-µm-thick sections using a bone cutting system (Exakt Norderstedt, Germany) and evaluated for the new bone formation at the periosteal and endosteal envelope under UV light.

**Statistical analysis**

The data are reported as the means ± standard deviations (SDs). Differences were tested for significance using 3-factor repeated-measures analysis of variance (ANOVA) with interactions (SigmaPlot, version 12.2, Systat Software, Chicago, IL, USA). *Post hoc* comparisons of means with a Bonferroni correction for multiple comparisons were performed only when interaction effects were significant. *P* values less than 0.05 were considered statistically significant.

**Results**

**Animals**

All of the rats enrolled in the study showed a 15% increase in body weight regardless of their treatment group. Neither the surgery nor treatment had an effect on animal health.

**Dynamic weight bearing**

The MM rats showed a different pattern of weight distribution compared with the sham controls. The weight-bearing load on the front feet of the MM rats was approximately 20% greater relative to sham controls at the 5-week time point, while both sham and MM rats showed similar weight bearing on the front feet at the end of study. The load-bearing capacity of the operated limbs of the MM rats did not increase despite the gain in body weight. The weight-bearing loads placed on the left hind leg were only moderately increased at 5 weeks in all 3 groups of MM rats regardless of treatment and stayed similar in all study groups throughout the 10-week study (Figure 1).

**EPIC µCT evaluation of the articular cartilage at the medial tibial plateau**

The articular cartilage was significantly thicker in Zone 1, which borders the osteophytes, in all three groups of rats that received the MM surgery compared with the sham controls. The cartilage was either considerably thinner or completely missing in the lateral part of Zone 1 and in the entirety of Zone 2 in all 3 groups of MM rats compared with the sham controls Figure 2 A-D.

**Histology evaluation of the articular cartilage**

In the control sham rats, the articular cartilage at the medial tibial plateau grew progressively thicker from the most medial zone 1 to the most lateral part zone 3 (Figure 3A). The mechanical imbalance induced by the MM surgery resulted in cartilage thickening at the most medial half of zone 1, next to the osteophytes, whereas the cartilage in zone 2 was thin and or completely missing in all MM rats (Figure 3B-D).
Cortical bone geometry at the femoral mid-diaphysis

The µCT results confirmed that the left and right femurs of the sham rats had similar cortical bone properties at the femoral diaphysis. The left femurs of the MM rats showed cortical bone geometry similar to that of the left femurs of the sham rats. However, the right femurs of the MM rats showed lower values for cortical bone properties compared with the right femurs of the sham rats but also exhibited somewhat lower values for all measured parameters compared with the contralateral left femurs. Both femurs from zoledronate and PTH treated rats showed slightly higher values for cortical bone parameters relative to the MM controls. The left femurs of the rats treated with zoledronate and PTH showed somewhat higher values for cortical bone properties compared to corresponding right femurs (Table 1).
Table 1: Table 1 shows the μCT analysis of the cortical bone at the femoral mid-diaphysis (midshaft) for both the left (L) and right (R) femurs. The following parameters were evaluated: bone mineral density (BMD; mg/mm²), tissue volume (T. Volume; mm³), bone volume (B. Volume; mm³), bone marrow volume (B. M. Volume; mm³), cortical thickness (C. Thickness; mm), bone area (B. Area; mm²), polar moment of inertia (pMoI; mm⁴), maximal I value (I_max; mm²), minimal I value (I_min; mm²), and I_max/C_max (mm²).

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Unit</th>
<th>Leg</th>
<th>Sham (mm²)</th>
<th>MM (mm²)</th>
<th>MM/Zol (mm²)</th>
<th>MM/PTH (mm²)</th>
</tr>
</thead>
<tbody>
<tr>
<td>BMD</td>
<td>mg/mm²</td>
<td>L</td>
<td>1367.54 ± 23.23</td>
<td>1371.21 ± 20.51</td>
<td>1389.00 ± 22.20</td>
<td>1356.92 ± 23.25</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>1342.94 ± 16.81</td>
<td>1332.36 ± 9.62</td>
<td>1338.04 ± 11.67</td>
<td>1329.47 ± 13.20</td>
</tr>
<tr>
<td>T. Volume</td>
<td>mm³</td>
<td>L</td>
<td>6.38 ± 0.44</td>
<td>6.08 ± 0.37</td>
<td>6.27 ± 0.32</td>
<td>6.40 ± 0.29</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>6.37 ± 0.40</td>
<td>6.04 ± 0.55</td>
<td>6.05 ± 0.43</td>
<td>6.18 ± 0.32</td>
</tr>
<tr>
<td>B. Volume</td>
<td>mm³</td>
<td>L</td>
<td>3.05 ± 0.15</td>
<td>2.93 ± 0.10</td>
<td>3.06 ± 0.17</td>
<td>3.15 ± 0.14</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>2.99 ± 0.19</td>
<td>2.89 ± 0.23</td>
<td>2.93 ± 0.17</td>
<td>3.04 ± 0.13</td>
</tr>
<tr>
<td>BM. Volume</td>
<td>mm³</td>
<td>L</td>
<td>3.13 ± 0.31</td>
<td>3.15 ± 0.28</td>
<td>3.21 ± 0.19</td>
<td>3.25 ± 0.17</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>2.98 ± 0.23</td>
<td>3.12 ± 0.34</td>
<td>3.13 ± 0.28</td>
<td>3.14 ± 0.21</td>
</tr>
<tr>
<td>Co. Th</td>
<td>mm</td>
<td>L</td>
<td>0.79 ± 0.05</td>
<td>0.77 ± 0.01</td>
<td>0.81 ± 0.03a</td>
<td>0.82 ± 0.02a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>0.79 ± 0.03</td>
<td>0.75 ± 0.01</td>
<td>0.78 ± 0.02a</td>
<td>0.80 ± 0.01a</td>
</tr>
<tr>
<td>B. Area</td>
<td>mm³</td>
<td>L</td>
<td>7.24 ± 0.37</td>
<td>6.96 ± 0.24</td>
<td>7.25 ± 0.21a</td>
<td>7.47 ± 0.34a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>7.19 ± 0.26</td>
<td>6.71 ± 0.22</td>
<td>6.95 ± 0.21a</td>
<td>7.21 ± 0.30a</td>
</tr>
<tr>
<td>PMol</td>
<td>mm³</td>
<td>L</td>
<td>16.36 ± 1.60</td>
<td>15.46 ± 1.41a</td>
<td>16.21 ± 1.70a</td>
<td>17.15 ± 1.69a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>16.28 ± 1.13</td>
<td>14.40 ± 1.14a</td>
<td>14.95 ± 1.72</td>
<td>16.01 ± 1.58</td>
</tr>
<tr>
<td>I_max</td>
<td>mm²</td>
<td>L</td>
<td>10.18 ± 1.11</td>
<td>9.71 ± 0.98a</td>
<td>10.21 ± 0.94a</td>
<td>10.58 ± 1.21a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>10.18 ± 0.68</td>
<td>8.79 ± 0.70a</td>
<td>9.26 ± 1.08</td>
<td>9.68 ± 1.02</td>
</tr>
<tr>
<td>I_min</td>
<td>mm²</td>
<td>L</td>
<td>6.18 ± 0.54</td>
<td>5.75 ± 0.49</td>
<td>5.99 ± 0.85</td>
<td>6.57 ± 0.54a</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>6.17 ± 0.53</td>
<td>5.61 ± 0.53</td>
<td>5.70 ± 0.72</td>
<td>6.34 ± 0.65a</td>
</tr>
<tr>
<td>I_max/C_max</td>
<td>mm²</td>
<td>L</td>
<td>4.53 ± 0.35</td>
<td>4.38 ± 0.28</td>
<td>4.58 ± 0.34</td>
<td>4.67 ± 0.35</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R</td>
<td>4.45 ± 0.22</td>
<td>4.10 ± 0.24</td>
<td>4.30 ± 0.40</td>
<td>4.38 ± 0.30</td>
</tr>
</tbody>
</table>

p<0.05 or p<0.01 relative to the sham rats. *p<0.05 or **p<0.01 relative to the MM rats.

Cortical bone strength

The left femurs of all 4 study groups showed similar maximum load parameters. The right femurs of the rats that underwent MM surgery displayed significantly lower maximum loads and energy to break values compared with the contralateral left femurs of the same study group and the right femurs of the sham rats (Figure 4).

Dynamic histomorphometry of the cortical bone

The rats in the sham group showed a similar intensity of bone mineralization in both the left and right femurs and at both cortical envelopes. The rats that underwent MM surgery showed less intensive mineralization of the right (operated) legs compared with the contralateral left legs of the same dosing group. Additionally, the rats treated with PTH showed the most intense mineralization at both cortical envelopes while the zoledronate-treated rats showed minimal bone formation at the endosteal envelope in both femurs (Figure 5).

Discussion

Animal disease models for testing novel osteoarthritis therapies are the gold standard in the preclinical phase of drug development. Despite improvements in animal experimentation, technological advances and biomarkers, too many unnecessary and poorly designed animal studies are still being conducted, yielding redundant and misleading results that have not been clinically confirmed. Although the overall benefit of such studies to the medical community is negligible, the lack of standardized animal models, study designs and defined endpoints continues to enable the publishing of pointless data.

The recent study conducted by our group was one of a very few publications to show the lack of efficacy of antiresorptive and anabolic therapy on the key indicators of OA in a preclinical model of PTOA [24]. As summarized in this manuscript, study results reveal that neither zoledronate nor PTH had a substantial effect on cartilage deterioration, osteophyte formation, osteosclerosis and joint functionality. Similar to our findings, the clinical data fail to demonstrate a clear benefit of anabolic and antiresorptive therapies in OA patients [21,22] despite decades of the use of these agents to treat patients with skeletal maladies [32,33]. The animal models of bone diseases and the biomarkers of bone metabolism, mass and structure are highly predictive of clinical outcomes [34,35]. Similarly, animal models of OA also seem to closely replicate the pathophysiology of human disease and are endorsed by the OARSI governing body [23]. Therefore, the discrepancy between the preclinical results and the clinical findings regarding benefits of bisphosphonates and PTH

Citation: Bagi CM, Benyman E, Chang-Ning Liu, Bagi IB, Zakur DE, et al. (2016) Antiresorptive and Anabolic Bone Therapy Does not Improve Weight Bearing Capacity and Bone Strength in OA Rats. Peertechz J Orthop Rheumatol 1(1): 003-011. DOI: http://dx.doi.org/10.17352/pjor.000002
to treat OA require further evaluation. Based on the literature, we hypothesized that both therapies would maintain or improve the strength of the cortical bone in the OA limb and that the bones would thereby remain fit to withstand mechanical loads and provide a foundation implementing physical therapy. The published literature unequivocally indicates that therapy with PTH and bisphosphonates increases cortical bone mass and strength during physiologic loading and that the addition of physical exercise further promotes the drugs’ effects on the cortical bone [36,37]. Contrariwise, in animal models with decreased load-bearing activity, anabolic and antiresorptive therapy seems to be less effective [38,39], proving the “mechanostat” theory that bone mass and architecture are regulated in response to the local strains generated in the local tissue by functional loading [40,41]. The results of this study show that partial disuse and reduced weight-bearing loads negatively impact the cortical bone geometry of the operated leg, as all 3 groups of osteoarthritic rats consistently exhibited somewhat lower values for cortical bone denominators compared with the contralateral left leg. Even though decline in cortical bone qualities of the operated leg was rather small and reached the significant difference only for pMoI and I_max parameters, the change seems to be sufficient to significantly affect cortical bone strength. Also, sedentary life style of rats in laboratory condition and different biomechanics in quadrupeds can minimize the change in cortical bone remodeling imposed by diminished capacity of osteoarthritic leg to withstand the mechanical loads and that should be taken into account when extrapolating these data to human subjects.

Similar to our findings, previous studies have shown that injured, osteoarthritic legs have a diminished capacity to withstand mechanical loads [42,43]. Here, we demonstrate that even moderate disuse over long periods has a deteriorating effect on cortical bone energy to break and maximal loads and that neither anabolic nor antiresorptive treatment can fully compensate for lack of physiological weight-bearing loads. Although cortical bone geometry and stiffness (the ability of bone to resist deformation under a given load) is preserved with PTH and, to a lesser extent, with zoledronate, those therapies fail to significantly improve bone strength, perhaps because of impaired mineralization and changes in the material properties of the cortical bone [44].

The mechanical loading that occurs with weight-bearing exercise is a potent anabolic stimulus of muscle, bone, and cartilage and is
The data from this study led us to conclude that patients with acute traumatic cartilage injury will benefit most from the early implementation of physical therapy and moderate exercise [45-48], which should provide the optimal milieu for simultaneous treatment with other therapies (anti-inflammatory, regenerative, nutrient and vitamin, and bone therapies) to support healing of the articular cartilage and subchondral bone end to ensure that the mechanical functions of the injured joint are restored.

In recent years, the regulatory and scientific community has imposed stringent rules to ensure that the well-being of laboratory animals and that the 3Rs paradigm is implemented [25,49]. It has been estimated that 25 million animals are used in research every year [50]. To further improve the usefulness of animal studies investigators should carefully select a suitable disease model and design the study of appropriate duration taking into account the slow metabolism of the cartilage and subchondral bone to allow the complete assessment of joint functionality. Additionally, several independent methods should be mandatory for every in vivo study to evaluate key attributes of cartilage and bone physiology, including imaging and histological evaluation of cartilage and bone, histochemistry and serum biomarkers. The lack of appropriate skills, technologies or funding should not be an excuse to compromise and run incomplete and poorly designed studies. Publishing “negative” data, even in the form of summary, should be encouraged to further promote information sharing and deter scientists from conducting unnecessary and redundant work. To that end, scientific journals and conferences should enforce stricter rules and guidelines for publishing data from in vivo studies to ensure scientific excellence and the proper use of laboratory animals.

Conclusions

The various methods utilized in this study showed that aggressive treatment with zoledronate and PTH was not sufficient to prevent or correct the deterioration of the hyaline cartilage, osteocyte formation and the mechanical incapacity of the osteoarthritic knee. Additionally, partial disuse of the osteoarthritic leg over long periods weakens cortical bone strength, and neither anabolic nor antiresorptive treatment could compensate for the lack of physiological weight-bearing loads. Our data suggest that the quick restoration of the mechanical function of the injured knee is a mandatory first step to prevent cartilage deterioration and support the recovery of joint structures and that a multi-therapeutic approach is warranted to treat various aspects of post-traumatic OA.

Role of Funding Source

This study was supported by Pfizer Consumer Healthcare. The study’s sponsor had no role in the study design, data collection, and data analysis and data interpretation and did not participate in the manuscript’s writing or submission.

Competing Interest

All Pfizer authors are regular employees and have no conflicts of interest to declare. Isabela Bagi was a summer intern at Pfizer and has no conflict of interest to declare.

References


