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Review Article

How the histamine N-methyltransferase inhibitor metoprine alleviates methamphetamine reward

Abstract

Several agents that activate brain histaminergic neurotransmission have been reported to improve methamphetamine (METH)-induced behavioral aberrations. In this review, we present research demonstrating that pretreatment with metoprine, a selective inhibitor of histamine N-methyltransferase (HMT), attenuates the reinforcing effects of METH in mice. Pretreatment with metoprine decreased METH-induced reinforcement as evaluated in the conditioned place preference (CPP) test. Metoprine pretreatment alone produced an increase in the CPP score with the same score level as that observed in mice treated with METH plus metoprine. No changes in alternation behaviors or numbers of marbles buried were observed in metoprine-treated mice measured in the Y-maze test and in the marble burying test, respectively. The locomotor activity was augmented after metoprine administration. These observations suggest that metoprine alleviates METH-induced rewarding property and hyperlocomotion. Metoprine is likely to augment spontaneous locomotor activities without mood alterations or short-term memory impairment. Brain histaminergic system is a new hope for treatment of METH dependence.

effective treatments for METH dependence are needed. Unfortunately, various attempts at pharmacotherapy trials have yielded unpromising and inconsistent results with medications developed to date [15-17]. Several novel alternative approaches have been a matter of intense investigation more recently, including dopamine D₃ receptor antagonists and partial agonists [18], and manipulations of brain histamine systems [19]. The later possibility is addressed in this report.

Based on a variety of findings, an increasing amount of scientific attention has been focused recently on the histaminergic system with respect to its potential roles in the causes and treatment of psychotic disorders, drug addiction/abuse, and other psychiatric conditions. Content of the histamine metabolite tele-methylhistamine is increased in the cerebrospinal fluid of schizophrenic patients as compared with healthy control subjects [20]. Dysfunction of the histamine-forming enzyme histidine decarboxylase (HDC) has been suggested to contribute to Tourette's syndrome in humans, with similar phenomena observed in mouse models [21]. Moreover, based upon the positioning of histamine H₃ receptors in neural circuits influencing dopaminergic and striatal function, histamine H₃ receptor antagonists have been investigated as potential anti-psychotic drugs [22-24]. In addition, brain histaminergic neurons have been suggested to influence amphetamine reinforcement, which was reduced by

Introduction

Methamphetamine (METH) is a highly addictive psychomotor stimulant drug that is abused worldwide [1]. METH abuse results in numerous adverse effects after acute administration, as well as an array of adverse outcomes associated with binge use, long-term use, and withdrawal [2-4]. Acutely METH releases dopamine from synaptic terminals through multiple actions that include inducing reverse transport of dopamine via the dopamine transporter (DAT), impairing the function of the vesicular monoamine transporter-2 (VMAT2), leading to increased cytoplasmic dopamine concentrations, and inhibition of monoamine oxidase [5-8]. Moreover, these changes contribute to the production of oxidative metabolites, metabolic impairments, oxidative damage to dopamine terminals, and depletion of tissue dopamine levels [9-11]. METH and related drugs consequently produce broad effects on the central nervous system both acutely and chronically [12-14].

Due to its highly addictive properties, and the adverse consequences associated with acute and chronic use of METH,

lesion of histaminergic neurons [25]. In this review, we will examine other recent evidence suggesting that histamine may have a role in drug reinforcement and addiction. The work that will be discussed will deal mainly with the relationship between alterations in brain histamine content and METH-induced behavior in animal models.

Brain histaminergic system

In the body histamine is synthesized by the decarboxylation of the amino acid L-histidine in a reaction catalyzed by HDC (EC4.1.1.22) and is stored mainly in mast cells, basophils, and neurons. Although a great deal of the histamine content of the brain comes from mast cells, histaminergic neurons in the brain are found solely in the tuberomammillary nucleus located in the posterior hypothalamus, but send projections throughout much of the central nervous system [26–28]. Regarding the function of histamine as a neurotransmitter, there are several important aspects of histamine neurotransmission that are important for understanding the role of histamine in brain function. (1) Aspects of histamine release are different from other monoamines. Depolarization releases neuronal histamine in the same fashion as other monoamines (i.e. dopamine, norepinephrine, and serotonin) [29–31], but the levels of extracellular histamine released are at most twice basal levels [30,31]. In contrast to histamine, other monoamines reach extracellular levels after neuronal depolarization-dependent releases that are several hundred-fold basal levels. (2) Aspects of histamine transport are different from other monoamines. There is no evidence for “histamine specific transporter” responsible for histamine clearance [32], as is the case for the other monoamine transporters, although histamine levels in the synaptic cleft return to a basal level after stimulation [33,34]. The organic cation transporter 3 (SLC22A3), previously called the extraneuronal monoamine transporter, is believed to maintain tissue histamine levels [35–37]. Concentrations of monoamines are regulated by SLC6 type high-affinity monoamine transporters [38], including DAT (SLC6A3) [39–41], the serotonin transporter (SERT; SLC6A4) [42–44], and the norepinephrine transporter (NET; SLC6A2) [45,46]. Despite current limitations in our knowledge of brain histamine dynamics, histamine is believed to be released from presynaptic vesicles by stimulation, and bind to histamine receptors located on postsynaptic (subtypes H_1 - H_4) and presynaptic (mainly H_3) membranes [47–50]. Histamine receptors are thought to have crucial roles in many physiological functions, including the sleep-wake cycle, food intake, neuroendocrine regulation, cognition, and drug reinforcement [51–54]. Histaminergic neurotransmission is terminated by metabolic inactivation of histamine by a histamine degrading enzyme histamine N-methyltransferase (HMT; EC2.1.1.8) [55]. Both HMT mRNAs and HMT protein-like immunoreactivity are located predominantly in the central nervous system [56–58]. HMT is considered to be the primary mechanism of histamine metabolism in brain, while histamine is metabolized by diamine oxidase (histaminase; EC1.4.3.6) in peripheral tissues [47,59]. Metoprine (2,4-diamino-5-(3',4'-dichlorophenyl)-6-methylpyrimidine) is an HMT inhibitor [60], which was originally developed as an anticancer drug

[61]. Metoprine easily crosses the blood-brain barrier when administered systemically [62] and increases tissue histamine content [63–68]. The involvement of central histaminergic systems in the behavioral and psychological effects of METH dependence can thus be investigated in using metoprine.

METH-induced behavior and metoprine

Acute METH administration releases histamine in the hypothalamus [69], and increases tissue histamine content [70]. The physiological relevance of METH-induced histamine release is unknown, but two possibilities might account for the significance of histamine release. Firstly, histamine released by METH might be associated with METH-induced behavior. This would be the most obvious view of the relationship between METH and histamine function. However, some evidence suggests otherwise. For instance, in histamine-deficient mice METH-induced hyperlocomotion and behavioral sensitization are augmented as compared with wild-type mice [71]. Alternatively, it might be suggested that histamine release is a part of a compensatory mechanism that restrains METH behavioral responses, limiting METH effects and the development of some chronic adaptations to METH such as sensitization. A failure of homeostasis in the brain resulting from reduced METH-induced histamine release may contribute to development of aberrant behaviors associated with chronic METH administration. If this is the case, it might be possible that these behaviors might be improved by increasing brain histamine levels pharmacologically.

There is some evidence, that high-dose METH-induced behaviors are reduced by agents which increase brain histamine content. L-histidine, a precursor for histamine synthesis, crosses the blood-brain barrier with a low K_m value [72] and is converted to histamine by HDC in the brain, increasing tissue histamine content [67,73]. Pretreatment with high doses of L-histidine attenuates METH-induced hyperlocomotion [64], behavioral sensitization [74,75], and stereotyped behavior [73,76,77]. The HMT inhibitors metoprine and the dimaprit analogue SKF 91488 (*S*-[4-(*N,N*-dimethylamino)butyl] isothioureia) increase tissue histamine content by inhibiting HMT [63–68,78], and these agents attenuate METH-induced stereotypy [63]. In line with these observations, it is likely that the increase in the tissue histamine contents might improve aberrant behavior observed at high METH doses, as exemplified by stereotypy and sensitization. This hypothesis is supported by evidence that METH-induced behaviors are exacerbated when histaminergic neurotransmission is suppressed by pharmacological [70] or genetic [71] manipulations.

Although this evidence supports a role for histamine in counteracting some of the behaviors observed after administration of high METH doses, it has not been determined whether effects observed at low METH doses, such as the rewarding and reinforcing effects, are similarly affected. To address this question, we examined the effect of metoprine pretreatment on METH-induced CPP in mice. In CPP testing, mice are initially presented with a testing apparatus that has two or three distinctive compartments (distinctive in terms of visual and tactile cues). Mice are allowed to explore

the compartments and the preference to each department is determined over 1–3 sessions. Ideally the preference for the compartments is approximately equal, although pairing with the less-preferred side is also effective (see review by Tzschentke [79] for a discussion of this issue). Subsequently, mice are confined to one of the compartments and receive an injection of a test drug (such as METH) in one compartment and saline in another compartment. After a series of such pairings over several days, preference is assessed again. An increase in preference reflects the positively reinforcing effects of the drugs, indicative of drug reward, while a decrease in preference reflects avoidance, indicative of aversive drug effects. Using this procedure mice readily develop a conditioned place preference for METH after 3 pairings with 0.5 mg/kg METH i.p. [80]. During the test locomotor stimulant effects of METH can also be measured.

Metoprine alone can produce locomotor stimulation, depending on dose [63,66], and thus might also be speculated to have some reinforcing effects when administered alone. In order to look at the potential interactive effects of modulating histamine with metoprine on METH reinforcement, mice were first examined in a modified conditioning procedure. Mice received only a single injection of METH (0.5 mg/kg, i.p.) during a single conditioning session (and one saline injection in the opposite compartment in a second session). Mice were then given a CPP test the following day, followed by a second test 5 days later. Control mice received saline injections during both training sessions. As shown in (Figure 1A), even a single METH injection produced significant CPP. This preference was reduced, but still significant, 5 days later. A second group of mice were pretreated with metoprine (10 mg/kg, i.p.) or saline, 1 h before METH conditioning (again control subjects received saline injections). The main finding of this study is that a pretreatment with metoprine reduced METH-induced CPP (Figure 1B). The result suggests that behavioral effects produced by relatively low doses of METH are reduced by activation of brain histaminergic systems. However, at the same time metoprine induced CPP when administered to control mice, suggesting that it may have reinforcing effects

of its own. Nonetheless, the data is consistent with a potential inhibitory effect of brain histaminergic activation, as shown by metoprine pretreatment on METH-induced CPP. In any case further investigation is warranted.

The extent to which such effects may represent specific effects of histaminergic modulation of METH reinforcement, or whether histaminergic modulation may affect drug reinforcement more generally, is also open to question. At least some data suggests that there may be broader effects of histamine modulation on drug reinforcement. The H_1 receptor antagonist chlorpheniramine can produce leftward shifts in the dose response curves for METH and cocaine CPP [81], although it must be noted that this drug also has effects on NET and SERT [82]. However, more selective histamine antagonists have been shown to affect morphine antinociception [83], conditioned place preference [84], and drug discrimination [85,86]. Moreover, L-histidine attenuated, while a histidine decarboxylase inhibitor potentiated, morphine CPP [84], similar to the effects observed for METH that were discussed previously. In non-human primates histamine antagonists can also be reinforcing alone or in combination with other drugs of abuse [87–91], although this may not be the case for selective H_3 antagonists [92]. Indeed, selective H_3 antagonists have been shown to reduce reinstatement of ethanol-seeking behavior during a reinstatement procedure after responding has been extinguished [93]. However, H_3 antagonists have also been reported to potentiate METH self-administration and METH-stimulated dopamine release [94], so the role of this receptor in different aspects of drug reinforcement, as well as other effects of drugs of abuse, is not yet clear.

Other metoprine effects

As mentioned above, mice and rats exhibit hyperlocomotion after metoprine administration [63,66], and these effects are dose-dependent [63]. This hyperlocomotion exhibits a bell-shaped dose-response curve, typical of many psychostimulant drugs. This is also similar to the effects of exogenously administered histamine on hyperlocomotion [95], effects thought to involve modulation of striatal dopamine function. This dose-response relationship suggests that different effects of elevating histamine levels emerge at low and high doses. The nature of these effects remains to be fully elucidated.

Other data suggests that there may be additional effects of metoprine-mediated histamine stimulation, on other spontaneous behaviors that might influence measures of drug reinforcement or hyperlocomotion, including perhaps effects on cognition. Histaminergic modulation is well-established to influence aspects of cognition [96,97], although much of this work is based upon studies with histamine antagonists. With regard to metoprine, however, in the Y-maze the number of spontaneous alternations were not different after treatment with metoprine (10 mg/kg) administration (Figure 2A), suggesting that metoprine did not affect the memory functions associated with this test. This behavior was unaffected by metoprine despite the presence of hyperlocomotion. This locomotion occurred even after administration of pyrilamine (10 mg/kg),

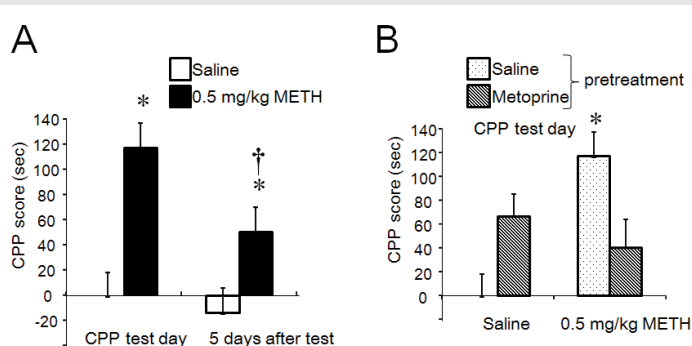


Figure 1: (A) A single injection of METH (0.5 mg/kg, i.p.) induces CPP. CPP was significant even 5 days after the CPP test day. Values are shown as the mean \pm SEM ($n = 12$). * $P < 0.05$, compared with saline/saline-conditioned mice; † $P < 0.05$, compared with CPP test day. (B) Effects of metoprine (10 mg/kg, i.p.) pretreatment on METH-induced CPP. Values are shown as the mean \pm SEM ($n = 12$). * $P < 0.05$, compared with saline/saline-treated mice; † $P < 0.05$, compared with METH/saline-treated mice (post hoc Bonferroni/Dunn test). METH, methamphetamine.

a histamine H_1 receptor antagonist (Figure 2B), suggesting that released histamine after metoprine administration might bind to histamine receptor subtypes other than the H_1 subtype. Metoprine is reported to exhibit an anxiogenic-like effect in mice as measured by the light/dark box test [98], although there are contradictory results as measured by the elevated plus maze test [63]. However, in the marble burying test, we found that metoprine did not have an anxiolytic-like profile (Figure 3). In addition, metoprine did not induce aggressive biting behavior (Figure 4A) as evaluated in the Aggression Response Meter (ARM) described previously [99], but metoprine induced hyperlocomotion even inside the animal chamber of the ARM (Figure 4B). Although, certainly examination of metoprine effects in other tests of anxiety and cognition are warranted, these initial investigations suggest that metoprine is not likely to augment spontaneous locomotor activity due to effects on anxiety or short-term memory function. These initial findings are therefore encouraging in that they suggest that metoprine may not cause side effects or toxicity at behaviorally relevant

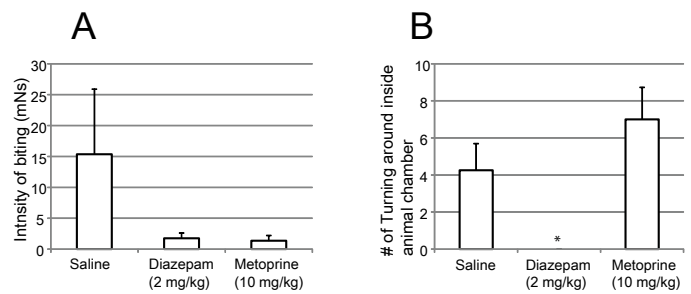


Figure 4: Intensity of aggressive biting (A) and number of turning around inside the animal chamber (B) after drug administration in mice. Values are shown as the mean \pm SEM ($n = 3$ or 4). * $P < 0.05$, compared with saline-treated mice (post hoc Bonferroni/Dunn test).

doses that would limit its utility as a potential approach to reducing METH-induced behaviors, which may potentially include those related to the reinforcing effects of METH, or those producing aberrant behavior at higher doses. This profile is further improved by the observation that HMT inhibitors activate central, but not peripheral, histaminergic systems so that these compounds are good candidates for evaluation as medications for the treatment of METH abuse and dependence.

HMT inhibitors as a new potential approach for the treatment of METH dependence

As argued in the preceding pages, modulation of histaminergic function may affect METH-induced behavior, including behavior relevant to METH abuse and METH dependence. In further support of this hypothesis, we have found that pretreatment with agmatine (decarboxylated L-arginine) attenuates METH-induced hyperlocomotion and stereotypy [100]. Agmatine is an endogenous cationic polyamine synthesized after decarboxylation of L-arginine by the enzyme arginine decarboxylase (EC4.1.1.19). As a possible neuromodulator in the brain, it binds to several receptors including the imidazoline I_1 , α_2 -adrenergic, and N-methyl-D-aspartate (NMDA) glutamate receptors [101-103]. However, of relevance to the present discussion, we have found that agmatine increases the tissue content of histamine in the hypothalamus (Figure 5A), but only in mice treated with METH. The histamine metabolite tele-methylhistamine was not affected (Figure 5B). Although the underlying mechanism is not clear at present, this data suggests that the inhibitory effect of agmatine on METH-induced hyperlocomotion and stereotypy may depend on the activation of the brain histaminergic systems.

Collectively, the data presented here supports the idea that augmentation of brain histamine content may limit behavioral effects of METH that may be relevant to METH abuse and dependence. This idea certainly needs further investigation, particularly given the, at times, inconsistent effects of histamine antagonists. However, one final encouraging finding is that deletion of histamine H_3 receptor genes attenuates METH-induced hyperlocomotion [104], consistent with our observations that elevating brain histamine function reduces METH effects [63,73], supporting the idea that activation of brain histamine systems may be a good strategy

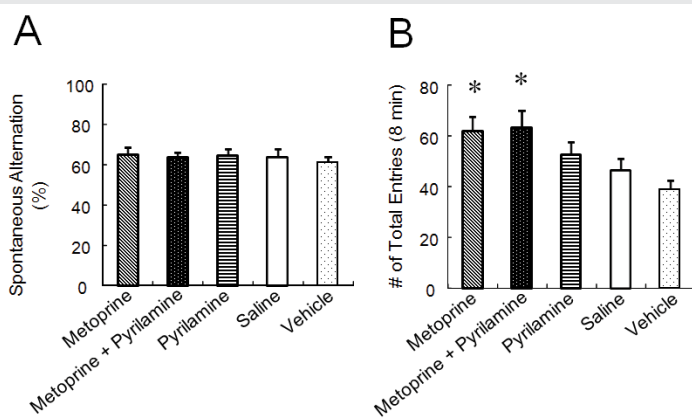


Figure 2: The effects of metoprine on spontaneous alternation behavior (A) and the total number of arm entries (B) in the Y-maze. Metoprine (and/or pyrilamine, a histamine H_1 receptor antagonist) was injected i.p. at a dose of 10 mg/kg. Values are shown as the mean \pm SEM ($n = 8$). * $P < 0.05$, compared with vehicle- and saline-treated mice (post hoc Bonferroni/Dunn test).

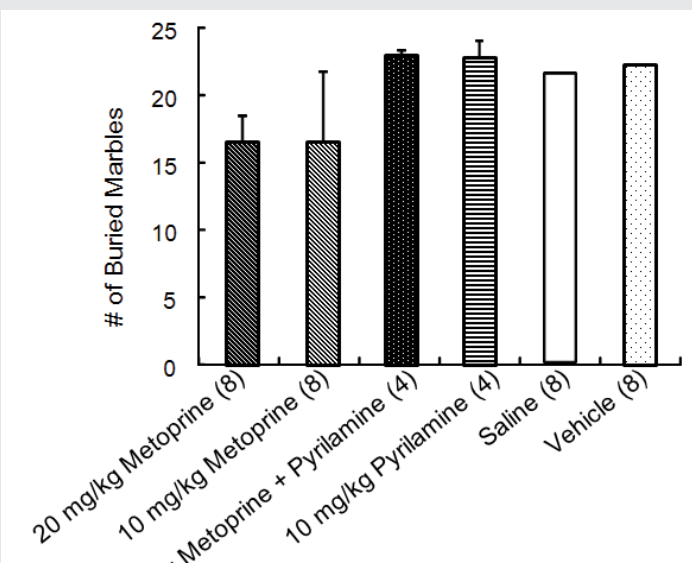


Figure 3: Effects of metoprine on marble burying behavior in mice. Values are shown as the mean \pm SEM ($n = 4$ or 8).

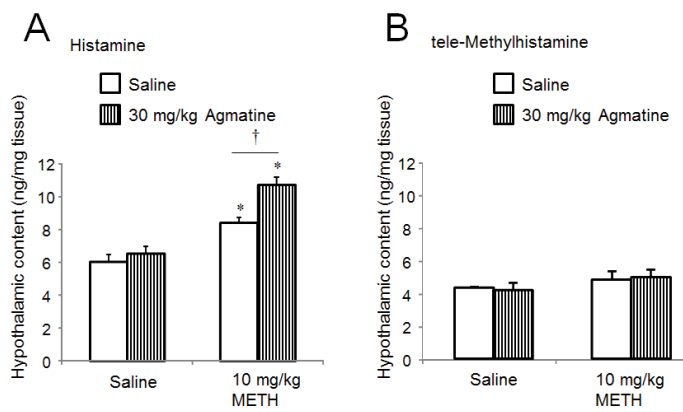


Figure 5: Hypothalamic content of histamine (A) and tele-methylhistamine (B). Mice were pretreated with agmatine (or saline) 1 h prior to METH (or saline). Brains were rapidly removed 1 h after METH treatment and hypothalamic regions were isolated. * $P < 0.05$, compared with saline-treated mice; † $P < 0.05$, compared with saline/METH-treated mice (post hoc Bonferroni/Dunn test). METH, methamphetamine.

for the development of agents which treat METH abuse and dependence. In line with these observations, applications of the HMT inhibitors like metoprine for routine clinical practice will offer additional insight into effective treatment for METH addiction and abuse.

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Conflicts of Interest

The authors declare no conflicts of interest associated with this manuscript.

References

- Courtney KE, Ray LA (2014) Methamphetamine: an update on epidemiology, pharmacology, clinical phenomenology, and treatment literature. *Drug and alcohol dependence* 143: 11-21. [Link: https://goo.gl/26Wwcy](https://goo.gl/26Wwcy)
- Harro J (2015) Neuropsychiatric Adverse Effects of Amphetamine and Methamphetamine. *Int Rev Neurobiol* 120: 179-204. [Link: https://goo.gl/JuPeo2](https://goo.gl/JuPeo2)
- Jablonski SA, Williams MT, Vorhees CV (2016) Neurobehavioral Effects from Developmental Methamphetamine Exposure. *Curr Top Behav Neurosci* 29: 183-230. [Link: https://goo.gl/7sSAH1](https://goo.gl/7sSAH1)
- Murray JB (1998) Psychophysiological aspects of amphetamine-methamphetamine abuse. *J Psychol* 132: 227-237. [Link: https://goo.gl/Tsjo5R](https://goo.gl/Tsjo5R)
- Panenka WJ, Procyshyn RM, Lecomte T, MacEwan GW, Flynn SW, et al. (2013) Methamphetamine use: a comprehensive review of molecular,

preclinical and clinical findings. *Drug Alcohol Depend* 129: 167-179. [Link: https://goo.gl/oTqtqz](https://goo.gl/oTqtqz)

- Sulzer D, Rayport S (1990) Amphetamine and other psychostimulants reduce pH gradients in midbrain dopaminergic neurons and chromaffin granules: a mechanism of action. *Neuron* 5: 797-808. [Link: https://goo.gl/NMMHxT](https://goo.gl/NMMHxT)
- Brown JM, Riddle EL, Sandoval V, Weston RK, Hanson JE, et al. (2002) A single methamphetamine administration rapidly decreases vesicular dopamine uptake. *J Pharmacol Exp Ther* 302: 497-501. [Link: https://goo.gl/YY5FaC](https://goo.gl/YY5FaC)
- Suzuki O, Hattori H, Asano M, Oya M, Katsumata Y (1980) Inhibition of monoamine oxidase by d-methamphetamine. *Biochemical pharmacology* 29: 2071-2073. [Link: https://goo.gl/qUs6FF](https://goo.gl/qUs6FF)
- Yamamoto BK, Zhu W (1998) The effects of methamphetamine on the production of free radicals and oxidative stress. *J Pharmacol Exp Ther* 287: 107-114. [Link: https://goo.gl/fa4845](https://goo.gl/fa4845)
- Schmidt CJ, Ritter JK, Sonsalla PK, Hanson GR, Gibb JW (1985) Role of dopamine in the neurotoxic effects of methamphetamine. *J Pharmacol Exp Ther* 233: 539-544. [Link: https://goo.gl/2V5xKS](https://goo.gl/2V5xKS)
- Jayanthi S, Ladenheim B, Cadet JL (1998) Methamphetamine-induced changes in antioxidant enzymes and lipid peroxidation in copper/zinc-superoxide dismutase transgenic mice. *Ann N Y Acad Sci* 844: 92-102. [Link: https://goo.gl/oM4UwT](https://goo.gl/oM4UwT)
- Bernheim A, See RE, Reichel CM (2016) Chronic methamphetamine self-administration disrupts cortical control of cognition. *Neuroscience and biobehavioral reviews* 69: 36-48. [Link: https://goo.gl/S1Y1k3](https://goo.gl/S1Y1k3)
- London ED, Kohno M, Morales AM, Ballard ME (2015) Chronic methamphetamine abuse and corticostriatal deficits revealed by neuroimaging. *Brain Res* 1628: 174-185. [Link: https://goo.gl/N7hIVC](https://goo.gl/N7hIVC)
- Northrop NA, Yamamoto BK (2015) Methamphetamine effects on blood-brain barrier structure and function. *Front Neurosci* 9: 69. [Link: https://goo.gl/gvTBHi](https://goo.gl/gvTBHi)
- Brensilver M, Heinzerling KG, Shoptaw S (2013) Pharmacotherapy of amphetamine-type stimulant dependence: an update. *Drug Alcohol Rev* 32: 449-460. [Link: https://goo.gl/5Merdx](https://goo.gl/5Merdx)
- Karila L, Weinstein A, Aubin HJ, Benyamina A, Reynaud M, et al. (2010) Pharmacological approaches to methamphetamine dependence: a focused review. *Br J Clin Pharmacol* 69: 578-592. [Link: https://goo.gl/UvkNjg](https://goo.gl/UvkNjg)
- Winslow BT, Voorhees KI, Pehl KA (2007) Methamphetamine abuse. *American family physician* 76: 1169-1174. [Link: https://goo.gl/kBM18b](https://goo.gl/kBM18b)
- Keck TM, John WS, Czoty PW, Nader MA, Newman AH (2015) Identifying Medication Targets for Psychostimulant Addiction: Unraveling the Dopamine D3 Receptor Hypothesis. *J Med Chem* 58: 5361-5380. [Link: https://goo.gl/ajjSvf](https://goo.gl/ajjSvf)
- Kitanaka J, Kitanaka N, Hall FS, Uhl GR, Takemura M (2016) Brain Histamine N-Methyltransferase As a Possible Target of Treatment for Methamphetamine Overdose. *Drug target insights* 10: 1-7. [Link: https://goo.gl/GhQybe](https://goo.gl/GhQybe)
- Prell GD, Green JP, Kaufmann CA, Khandelwal JK, Morrishow AM, et al. (1995) Histamine metabolites in cerebrospinal fluid of patients with chronic schizophrenia: their relationships to levels of other aminergic transmitters and ratings of symptoms. *Schizophr Res* 14: 93-104. [Link: https://goo.gl/2OkWHW](https://goo.gl/2OkWHW)
- Castellan Baldan L, Williams KA, Gallezot JD, Pogorelov V, Rapanelli M, et al. (2014) Histidine decarboxylase deficiency causes tourette syndrome: parallel findings in humans and mice. *Neuron* 81: 77-90. [Link: https://goo.gl/GtbsQ](https://goo.gl/GtbsQ)
- Arrang JM (2007) Histamine and schizophrenia. *Int Rev Neurobiol* 78: 247-287. [Link: https://goo.gl/N3HS3f](https://goo.gl/N3HS3f)
- Baronio D, Gonchoroski T, Castro K, Zanatta G, Gottfried C, et al. (2014)

- Histaminergic system in brain disorders: lessons from the translational approach and future perspectives. *Annals of general psychiatry* 13: 34. [Link: https://goo.gl/DH09Oq](https://goo.gl/DH09Oq)
24. Ellenbroek BA, Ghiabi B (2015) Do Histamine receptor 3 antagonists have a place in the therapy for schizophrenia? *Curr Pharm Des* 21: 3760-3770. [Link: https://goo.gl/7qaCqj](https://goo.gl/7qaCqj)
 25. Contreras M, Riveros ME, Quispe M, Sanchez C, Perdomo G, et al. (2016) The Histaminergic Tuberoamillary Nucleus Is Involved in Appetite for Sex, Water and Amphetamine. *PloS one* 11: e0148484. [Link: https://goo.gl/OfSwP1](https://goo.gl/OfSwP1)
 26. Panula P, Pirvola U, Auvinen S, Airaksinen MS (1989) Histamine-immunoreactive nerve fibers in the rat brain. *Neuroscience* 28: 585-610. [Link: https://goo.gl/saZZkQ](https://goo.gl/saZZkQ)
 27. Panula P, Yang HY, Costa E (1984) Histamine-containing neurons in the rat hypothalamus. *Proc Natl Acad Sci U S A* 81: 2572-2576. [Link: https://goo.gl/aDCey4](https://goo.gl/aDCey4)
 28. Watanabe T, Taguchi Y, Hayashi H, Tanaka J, Shiosaka S, et al. (1983) Evidence for the presence of a histaminergic neuron system in the rat brain: an immunohistochemical analysis. *Neuroscience letters* 39: 249-254. [Link: https://goo.gl/CBYoHe](https://goo.gl/CBYoHe)
 29. Arrang JM, Devaux B, Chodkiewicz JP, Schwartz JC (1988) H3-receptors control histamine release in human brain. *J Neurochem* 51: 105-108. [Link: https://goo.gl/BnEOHT](https://goo.gl/BnEOHT)
 30. Mochizuki T, Yamatodani A, Okakura K, Takemura M, Inagaki N, et al. (1991) In vivo release of neuronal histamine in the hypothalamus of rats measured by microdialysis. *Naunyn-Schmiedeberg's archives of pharmacology* 343: 190-195. [Link: https://goo.gl/ihNsZj](https://goo.gl/ihNsZj)
 31. Ono J, Yamatodani A, Kishino J, Okada S, Wada H (1992) Cholinergic influence of K(+)-evoked release of endogenous histamine from rat hypothalamic slices in vitro. *Methods Find Exp Clin Pharmacol* 14: 35-40. [Link: https://goo.gl/j4ub4K](https://goo.gl/j4ub4K)
 32. Ogasawara M, Yamauchi K, Satoh Y, Yamaji R, Inui K, et al. (2006) Recent advances in molecular pharmacology of the histamine systems: organic cation transporters as a histamine transporter and histamine metabolism. *J Pharmacol Sci* 101: 24-30. [Link: https://goo.gl/u4n3p2](https://goo.gl/u4n3p2)
 33. Okakura K, Yamatodani A, Mochizuki T, Horii A, Wada H (1992) Glutamatergic regulation of histamine release from rat hypothalamus. *Eur J Pharmacol* 213: 189-192. [Link: https://goo.gl/xEriY9](https://goo.gl/xEriY9)
 34. John J, Kodama T, Siegel JM (2014) Caffeine promotes glutamate and histamine release in the posterior hypothalamus. *Am J Physiol Regul Integr Comp Physiol* 307: R704-710. [Link: https://goo.gl/E4I0xy](https://goo.gl/E4I0xy)
 35. Yoshikawa T, Naganuma F, Iida T, Nakamura T, Harada R, et al. (2013) Molecular mechanism of histamine clearance by primary human astrocytes. *Glia* 61: 905-916. [Link: https://goo.gl/whzNpL](https://goo.gl/whzNpL)
 36. Schneider E, Machavoine F, Pléau JM, Bertron AF, Thurmond RL, et al. (2005) Organic cation transporter 3 modulates murine basophil functions by controlling intracellular histamine levels. *J Exp Med* 202: 387-393. [Link: https://goo.gl/96pKW1](https://goo.gl/96pKW1)
 37. Gasser PJ, Hurley MM, Chan J, Pickel VM (2016) Organic cation transporter 3 (OCT3) is localized to intracellular and surface membranes in select glial and neuronal cells within the basolateral amygdaloid complex of both rats and mice. *Brain Struct Funct* 222: 1913-1928. [Link: https://goo.gl/tchVxW](https://goo.gl/tchVxW)
 38. Pramod AB, Foster J, Carvelli L, Henry LK (2013) SLC6 transporters: structure, function, regulation, disease association and therapeutics. *Mol Aspects Med* 34: 197-219. [Link: https://goo.gl/ybzjXj](https://goo.gl/ybzjXj)
 39. Kristensen AS, Andersen J, Jørgensen TN, Sørensen L, Eriksen J (2011) SLC6 neurotransmitter transporters: structure, function, and regulation. *Pharmacological reviews* 63: 585-640. [Link: https://goo.gl/VgR7Tn](https://goo.gl/VgR7Tn)
 40. Lin Z, Canales JJ, Björgvinsson T, Thomsen M, Qu H, et al. (2011) Monoamine transporters: vulnerable and vital doorkeepers. *Prog Mol Biol Transl Sci* 98: 1-46. [Link: https://goo.gl/NUv7wJ](https://goo.gl/NUv7wJ)
 41. Giros B, el Mestikawy S, Bertrand L, Caron MG (1991) Cloning and functional characterization of a cocaine-sensitive dopamine transporter. *FEBS Lett* 295: 149-154. [Link: https://goo.gl/meYg3n](https://goo.gl/meYg3n)
 42. Lesch KP, Wolozin BL, Estler HC, Murphy DL, Riederer P (1993) Isolation of a cDNA encoding the human brain serotonin transporter. *J Neural Transm Gen Sect* 91: 67-72. [Link: https://goo.gl/BU65Gu](https://goo.gl/BU65Gu)
 43. Blakely RD, Berson HE, Fremeau RT Jr, Caron MG, Peek MM, et al. (1991) Cloning and expression of a functional serotonin transporter from rat brain. *Nature* 354: 66-70. [Link: https://goo.gl/wlo0n9](https://goo.gl/wlo0n9)
 44. Barr CS, Newman TK, Becker ML, Parker CC, Champoux M, et al. (2003) The utility of the non-human primate; model for studying gene by environment interactions in behavioral research. *Genes Brain Behav* 2: 336-340. [Link: https://goo.gl/So0p8f](https://goo.gl/So0p8f)
 45. Pacholczyk T, Blakely RD, Amara SG (1991) Expression cloning of a cocaine- and antidepressant-sensitive human noradrenaline transporter. *Nature* 350: 350-354. [Link: https://goo.gl/9V0ds4](https://goo.gl/9V0ds4)
 46. Apparsundaram S (2001) Function and regulation of monoamine transporters: focus on the norepinephrine transporter. *CNS Spectr* 6: 671-674. [Link: https://goo.gl/Xdn4o6](https://goo.gl/Xdn4o6)
 47. Haas HL, Sergeeva OA, Selbach O (2008) Histamine in the nervous system. *Physiol Rev* 88: 1183-1241. [Link: https://goo.gl/KmtEK5](https://goo.gl/KmtEK5)
 48. Prell GD, Green JP (1986) Histamine as a neuroregulator. *Annu Rev Neurosci* 9: 209-254. [Link: https://goo.gl/WPLQh5](https://goo.gl/WPLQh5)
 49. Timmerman H (1989) Histamine receptors in the central nervous system. *Pharm Weekbl Sci* 11: 146-150. [Link: https://goo.gl/WFrEJg](https://goo.gl/WFrEJg)
 50. Wada H, Inagaki N, Yamatodani A, Watanabe T (1991) Is the histaminergic neuron system a regulatory center for whole-brain activity? *Trends Neurosci* 14: 415-418. [Link: https://goo.gl/dBrxQL](https://goo.gl/dBrxQL)
 51. Alvarez EO (2009) The role of histamine on cognition. *Behav Brain Res* 199: 183-189. [Link: https://goo.gl/ZKXgy7](https://goo.gl/ZKXgy7)
 52. Brown RE, Stevens DR, Haas HL (2001) The physiology of brain histamine. *Prog Neurobiol* 63: 637-672. [Link: https://goo.gl/VUyWwP](https://goo.gl/VUyWwP)
 53. Knigge U, Warberg J (1991) Neuroendocrine functions of histamine. *Agents Actions Suppl* 33: 29-53. [Link: https://goo.gl/XDbhRR](https://goo.gl/XDbhRR)
 54. Brabant C, Alleva L, Quertemont E, Tirelli E (2010) Involvement of the brain histaminergic system in addiction and addiction-related behaviors: a comprehensive review with emphasis on the potential therapeutic use of histaminergic compounds in drug dependence. *Prog Neurobiol* 92: 421-441. [Link: https://goo.gl/0hQRuu](https://goo.gl/0hQRuu)
 55. Takemura M, Kitanaka N, Kitanaka J (2003) Signal transduction by histamine in the cerebellum and its modulation by N-methyltransferase. *Cerebellum* 2: 39-43. [Link: https://goo.gl/1nnbzY](https://goo.gl/1nnbzY)
 56. Takemura M, Imamura I, Mizuguchi H, Fukui H, Yamatodani A (1994) Tissue distribution of histamine N-methyltransferase-like immunoreactivity in rodents. *Life Sci* 54: 1059-1071. [Link: https://goo.gl/9Nvucv](https://goo.gl/9Nvucv)
 57. Kitanaka J, Kitanaka N, Tsujimura T, Kakahana M, Terada N, et al. (2001) Guinea pig histamine N-methyltransferase: cDNA cloning and mRNA distribution. *Japanese journal of pharmacology* 85: 105-108. [Link: https://goo.gl/loTyXD](https://goo.gl/loTyXD)
 58. Kitanaka N, Kitanaka J, Oue T, Tada Y, Tanaka T, et al. (2002) Genomic structure of the rat and mouse histamine N-methyltransferase gene. *Japanese journal of pharmacology* 88: 85-92. [Link: https://goo.gl/VEdPwZ](https://goo.gl/VEdPwZ)

59. Kitanaka J, Kitanaka N, Tsujimura T, Terada N, Takemura M (2002) Expression of diamine oxidase (histaminase) in guinea-pig tissues. *European journal of pharmacology* 437: 179-185. [Link: https://goo.gl/hTSSbZ](https://goo.gl/hTSSbZ)
60. Beaven MA, Shaff RE (1979) New inhibitors of histamine-N-methyltransferase. *Biochemical pharmacology* 28: 183-188. [Link: https://goo.gl/C9jPi1](https://goo.gl/C9jPi1)
61. Cavallito JC, Nichol CA, Brenckman WD Jr, Deangelis RL, Stickney DR, et al. (1978) Lipid-soluble inhibitors of dihydrofolate reductase. I. Kinetics, tissue distribution, and extent of metabolism of pyrimethamine, metoprine, and etoprine in the rat, dog, and man. *Drug Metab Dispos* 6: 329-337.
62. Nichol CA, Cavallito JC, Woolley JL, Sigel CW (1977) Lipid-soluble diaminopyrimidine inhibitors of dihydrofolate reductase. *Cancer treatment reports* 61: 559-564. [Link: https://goo.gl/OIXjCT](https://goo.gl/OIXjCT)
63. Kitanaka J, Kitanaka N, Tatsuta T, Morita Y, Takemura M (2007) Blockade of brain histamine metabolism alters methamphetamine-induced expression pattern of stereotypy in mice via histamine H₁ receptors. *Neuroscience* 147: 765-777. [Link: https://goo.gl/bJadbY](https://goo.gl/bJadbY)
64. Itoh Y, Nishibori M, Oishi R, Saeki K (1984) Neuronal histamine inhibits methamphetamine-induced locomotor hyperactivity in mice. *Neuroscience letters* 48: 305-309. [Link: https://goo.gl/jx6Sql](https://goo.gl/jx6Sql)
65. Sakai N, Onodera K, Maeyama K, Yanai K, Watanabe T (1992) Effects of (S)-alpha-fluoromethylhistidine and metoprine on locomotor activity and brain histamine content in mice. *Life sci* 51: 397-405. [Link: https://goo.gl/qCjBOE](https://goo.gl/qCjBOE)
66. Samotaeva IS, Birioukova LM, Midzyanovskaya IS, Kuznetsova GD, Bazyan AS, et al. (2012) Metoprine induced behavioral modifications and brain regional histamine increase in WAG/Rij and Wistar rats. *Epilepsy research* 101: 148-156. [Link: https://goo.gl/SxIQ11](https://goo.gl/SxIQ11)
67. Muroi N, Oishi R, Saeki K (1991) Effect of reserpine on histamine metabolism in the mouse brain. *J Pharmacol Exp Ther* 256: 967-972. [Link: https://goo.gl/waVaZb](https://goo.gl/waVaZb)
68. Zawilska J, Nowak JZ (1985) Changes in the rat brain histamine content following metoprine and other histamine-methyltransferase (HMT) inhibitors. *Pol J Pharmacol Pharm* 37: 821-830. [Link: https://goo.gl/DAH7d3](https://goo.gl/DAH7d3)
69. Ito C, Onodera K, Yamatodani A, Watanabe T, Sato M (1997) The effect of methamphetamine on histamine release in the rat hypothalamus. *Psychiatry and clinical neurosciences* 51: 79-81. [Link: https://goo.gl/TkADHx](https://goo.gl/TkADHx)
70. Kitanaka J, Kitanaka N, Hall FS, Uhl GR, Tatsuta T, et al. (2011) Histamine H₃ receptor agonists decrease hypothalamic histamine levels and increase stereotypical biting in mice challenged with methamphetamine. *Neurochem Res* 36: 1824-1833. [Link: https://goo.gl/jiK1Lb](https://goo.gl/jiK1Lb)
71. Kubota Y, Ito C, Sakurai E, Sakurai E, Watanabe T, et al. (2002) Increased methamphetamine-induced locomotor activity and behavioral sensitization in histamine-deficient mice. *J Neurochem* 83: 837-845. [Link: https://goo.gl/a8Pqne](https://goo.gl/a8Pqne)
72. Hargreaves KM, Pardridge WM (1988) Neutral amino acid transport at the human blood-brain barrier. *J Biol Chem* 263: 19392-19397. [Link: https://goo.gl/Ae146D](https://goo.gl/Ae146D)
73. Kitanaka J, Kitanaka N, Tatsuta T, Miyoshi A, Koumoto A, et al. (2010) Pretreatment with L-histidine produces a shift from methamphetamine-induced stereotypical biting to persistent locomotion in mice. *Pharmacol Biochem Behav* 94: 464-470. [Link: https://goo.gl/9df7tO](https://goo.gl/9df7tO)
74. Ito C, Onodera K, Watanabe T, Sato M (1997) Effects of histamine agents on methamphetamine-induced stereotyped behavior and behavioral sensitization in rats. *Psychopharmacology* 130: 362-367. [Link: https://goo.gl/9msARh](https://goo.gl/9msARh)
75. Ito C, Sato M, Onodera K, Watanabe T (1996) The role of the brain histaminergic neuron system in methamphetamine-induced behavioral sensitization in rats. *Ann N Y Acad Sci* 801: 353-360. [Link: https://goo.gl/fj2Dvc](https://goo.gl/fj2Dvc)
76. Joshi VV, Balsara JJ, Jadhav JH, Chandorkar AG (1981) Effect of L-histidine and chlorcyclizine on apomorphine-induced climbing behaviour and methamphetamine stereotypy in mice. *Eur J Pharmacol* 69: 499-502. [Link: https://goo.gl/NsRT7c](https://goo.gl/NsRT7c)
77. Muley MP, Balsara JJ, Chandorkar AG (1979) Effect of L-histidine pretreatment on methamphetamine induced stereotyped behaviour in rats. *Indian J Physiol Pharmacol* 23: 291-296. [Link: https://goo.gl/g9nK7p](https://goo.gl/g9nK7p)
78. Klein MC, Gertner SB (1981) Evidence for a role of endogenous histamine in central cardiovascular regulation: inhibition of histamine-N-methyltransferase by SKF 91488. *J Pharmacol Exp Ther* 216: 315-320. [Link: https://goo.gl/K3ySmB](https://goo.gl/K3ySmB)
79. Tzschentke TM (1998) Measuring reward with the conditioned place preference paradigm: a comprehensive review of drug effects, recent progress and new issues. *Prog Neurobiol* 56: 613-672. [Link: https://goo.gl/uH1ddY](https://goo.gl/uH1ddY)
80. Kitanaka N, Kitanaka J, Tatsuta T, Watabe K, Morita Y, et al. (2006) Methamphetamine reward in mice as assessed by conditioned place preference test with Supermex sensors: effect of subchronic clorgyline pretreatment. *Neurochem Res* 31: 805-813. [Link: https://goo.gl/DraRnT](https://goo.gl/DraRnT)
81. Masukawa Y, Suzuki T, Misawa M (1993) Differential modification of the rewarding effects of methamphetamine and cocaine by opioids and antihistamines. *Psychopharmacology* 111: 139-143. [Link: https://goo.gl/HuLH04](https://goo.gl/HuLH04)
82. Carlsson A, Lindqvist M (1969) Central and peripheral monoaminergic membrane-pump blockade by some addictive analgesics and antihistamines. *J Pharm Pharmacol* 21: 460-464. [Link: https://goo.gl/ZVnAOH](https://goo.gl/ZVnAOH)
83. Suzuki T, Takamori K, Takahashi Y, Narita M, Misawa M, et al. (1994) The differential effects of histamine receptor antagonists on morphine- and U-50,488H-induced antinociception in the mouse. *Life sci* 54: 203-211. [Link: https://goo.gl/H3Kfgt](https://goo.gl/H3Kfgt)
84. Suzuki T, Takamori K, Misawa M, Onodera K (1995) Effects of the histaminergic system on the morphine-induced conditioned place preference in mice. *Brain research* 675: 195-202. [Link: https://goo.gl/4CPTkR](https://goo.gl/4CPTkR)
85. Suzuki T, Mori T, Tsuji M, Misawa M, Onodera K (1997) Interactions between H₁-antagonists and opioids: a drug discrimination study. *Psychopharmacology* 131: 346-353. [Link: https://goo.gl/7hx13k](https://goo.gl/7hx13k)
86. Mori T, Narita M, Onodera K, Suzuki T (2004) Involvement of histaminergic system in the discriminative stimulus effects of morphine. *Eur J Pharmacol* 491: 169-172. [Link: https://goo.gl/z7CDKM](https://goo.gl/z7CDKM)
87. Banks ML, Andersen ML, Murnane KS, Meyer RC, Howell LL (2009) Behavioral and neurochemical effects of cocaine and diphenhydramine combinations in rhesus monkeys. *Psychopharmacology* 205: 467-474. [Link: https://goo.gl/3VjUNQ](https://goo.gl/3VjUNQ)
88. Beardsley PM, Balster RL (1992) The intravenous self-administration of antihistamines by rhesus monkeys. *Drug Alcohol Depend* 30: 117-126. [Link: https://goo.gl/3NnKJA](https://goo.gl/3NnKJA)
89. Bergman J, Speelman RD (1986) Some behavioral effects of histamine H₁ antagonists in squirrel monkeys. *J Pharmacol Exp Ther* 239: 104-110. [Link: https://goo.gl/oWftSb](https://goo.gl/oWftSb)
90. Wang Z, Woolverton WL (2007) Self-administration of cocaine-antihistamine combinations: super-additive reinforcing effects. *Eur J Pharmacol* 557: 159-160. [Link: https://goo.gl/VQQB4n](https://goo.gl/VQQB4n)
91. Wang Z, Woolverton WL (2009) Super-additive interaction of the reinforcing effects of cocaine and H₁-antihistamines in rhesus monkeys. *Pharmacol Biochem Behav* 91: 590-595. [Link: https://goo.gl/Dw16WB](https://goo.gl/Dw16WB)
92. Hudzik TJ, Basso A, Boyce-Rustay JM, Bracken W, Browman KE, et al. (2013) Assessment of the abuse liability of ABT-288, a novel histamine H(3) receptor antagonist. *Psychopharmacology* 228: 187-197. [Link: https://goo.gl/yQRIVT](https://goo.gl/yQRIVT)

93. Nuutinen S, Mäki T, Rozov S, Bäckström P, Hyytiä P, et al. (2016) Histamine H3 receptor antagonist decreases cue-induced alcohol reinstatement in mice. *Neuropharmacology* 106: 156-163. [Link: https://goo.gl/r3c7SJ](https://goo.gl/r3c7SJ)
94. Munzar P, Tanda G, Justinova Z, Goldberg SR (2004) Histamine h3 receptor antagonists potentiate methamphetamine self-administration and methamphetamine-induced accumbal dopamine release. *Neuropsychopharmacology* 29: 705-717. [Link: https://goo.gl/NzmOtc](https://goo.gl/NzmOtc)
95. Chiavegatto S, Nasello AG, Bernardi MM (1998) Histamine and spontaneous motor activity: biphasic changes, receptors involved and participation of the striatal dopamine system. *Life sci* 62: 1875-1888. [Link: https://goo.gl/GGyRdr](https://goo.gl/GGyRdr)
96. Van Ruitenbeek P, Vermeeren A, Riedel WJ (2010) Cognitive domains affected by histamine H(1)-antagonism in humans: a literature review. *Brain Res Rev* 64: 263-282. [Link: https://goo.gl/txScIz](https://goo.gl/txScIz)
97. Brioni JD, Esbenshade TA, Garrison TR, Bitner SR, Cowart MD (2011) Discovery of histamine H3 antagonists for the treatment of cognitive disorders and Alzheimer's disease. *J Pharmacol Exp Ther* 336: 38-46. [Link: https://goo.gl/3lf5ph](https://goo.gl/3lf5ph)
98. Malmberg-Aiello P, Ipponi A, Bartolini A, Schunack W (2002) Mouse light/dark box test reveals anxiogenic-like effects by activation of histamine H1 receptors. *Pharmacol Biochem Behav* 71: 313-318. [Link: https://goo.gl/8gTMDM](https://goo.gl/8gTMDM)
99. Kuchiiwa S, Kuchiiwa T (2014) A novel semi-automated apparatus for measurement of aggressive biting behavior in mice. *Journal of neuroscience methods* 228: 27-34. [Link: https://goo.gl/Siv5rD](https://goo.gl/Siv5rD)
100. Kitanaka N, Kitanaka J, Hall FS, Uhl GR, Watabe K, et al. (2014) Agmatine attenuates methamphetamine-induced hyperlocomotion and stereotyped behavior in mice. *Behav Pharmacol* 25: 158-165. [Link: https://goo.gl/6aCGcy](https://goo.gl/6aCGcy)
101. Halaris A, Plietz J (2007) Agmatine : metabolic pathway and spectrum of activity in brain. *CNS drugs* 21: 885-900. [Link: https://goo.gl/HWIPsG](https://goo.gl/HWIPsG)
102. Reis DJ, Regunathan S (2000) Is agmatine a novel neurotransmitter in brain? *Trends in pharmacological sciences* 21: 187-193. [Link: https://goo.gl/SxUtkY](https://goo.gl/SxUtkY)
103. Peters D, Berger J, Langnaese K, Derst C, Madai VI, et al. (2013) Arginase and Arginine Decarboxylase - Where Do the Putative Gate Keepers of Polyamine Synthesis Reside in Rat Brain? *PLoS one* 8: e66735. [Link: https://goo.gl/UU29Wx](https://goo.gl/UU29Wx)
104. Okuda T, Zhang D, Shao H, Okamura N, Takino N, et al. (2009) Methamphetamine- and 3,4-methylenedioxymethamphetamine-induced behavioral changes in histamine H3-receptor knockout mice. *J Pharmacol Sci* 111: 167-174. [Link: https://goo.gl/u85WyH](https://goo.gl/u85WyH)